Protein Analysis of Marjoram Plants Treated with Biocontrol Agents

Islam M. H. Rizk1*, Mohamed M. M. Amaar2 and Ibrahim A. Ibrahim3

1Department of Environmental Biotechnology, Genetic Engineering and Biotechnology Research Institute, University of Sadat City, Egypt.
2Department of Agricultural Botany, Faculty of Agriculture, Minufiya University, Egypt
3Department of Plant Biotechnology Genetic Engineering and Biotechnology Research Institute, University of Sadat City, Egypt.

*Corresponding Author; Email: islam_rizk2@yahoo.com

ABSTRACT

Origanum majorana L. commonly known as sweet marjoram has been used for variety of diseases in traditional medicine, including gastrointestinal, ocular, nasopharyngeal, respiratory, cardiac, rheumatologic, and neurological disorders. Protein gel electrophoresis is the commonly used technology to separate proteins according to their physical properties such as electrical charge and molecular weight. In present study, polyacrylamide gel electrophoresis (SDS-PAGE) was used for identification of marjoram plants treated with biocontrol agents collected from Egypt and studying the differentiations among them. SDS-PAGE was examined for plants treated with four biocontrol agents: Bacillus megaterium, Fusarium aqueductuum, Trichoderma harzinum + Fusarium aqueductuum and Bacillus megaterium + Trichoderma harzinum + Trichoderma hamatum + Fusarium aqueductuum. Protein analysis revealed that, application of the biocontrol agents significantly increased protein bands than control or plants cultivated in the infested soil, best results were obtained when B. megaterium was applied.

Keywords: Origanum majorana, traditional medicine, Fusarium aqueductuum, polyacrylamide gel electrophoresis (SDS-PAGE), Majorana hortensis

INTRODUCTION

Origanum majorana L. from the family Lamiaceae (syn. Majorana hortensis Moench) is commonly known as sweet marjoram. This herb is native to Mediterranean region and cultivated in many countries of Asia, North Africa, and Europe, for example, Spain, Hungary, Portugal, Germany, Egypt, Poland, and France. Origanum majorana grows up to 30 to 60 cm (Novak et al., 2008; El-Ghorab et al., 2004). It is a perennial bushy plant. It has oblique rhizome, hairy shrub like stalks, opposite dark green oval leaves and white or red flowers in clustered bracts. The leaves are whole, larger ones being fragmented, oblate to broadly elliptical. 1–3 This plant is widely used as a garnish and is used for different medicinal purposes in traditional and folklore medicine of different countries (Ramadan et al., 2013; Abdel-Massih et al., 2010). Various compounds have been identified in sweet marjoram. Also, different pharmacological activities have been attributed to this plant (Abd and El-Metwally, 2010; Al-Howiriny et al., 2009; Amarowicz et al., 2009). The present review summarizes comprehensive information concerning traditional uses, phytochemistry, and pharmacological activities of sweet marjoram. For this purpose,
databases, including PubMed, Google Scholar, and Scopus were searched for studies focusing on the ethnomedicinal use, phytochemical compounds and pharmacological activities of sweet marjoram. Data were collected from 1980 to 2015 (up to July). The search terms were “sweet marjoram” or “Origanum majorana” (Dragland et al., 2003)

Proteins are important constituents of foods for a number of different reasons. They are a major source of energy, as well as containing essential amino-acids, such as lysine, tryptophan, methionine, leucine, isoleucine and valine, which are essential to human health, but which the body cannot synthesize. Proteins are also the major structural components of many natural foods, often determining their overall texture, e.g., tenderness of meat or fish products. Isolated proteins are often used in foods as ingredients because of their unique functional properties, (i.e., their ability to provide desirable appearance, texture or stability). Typically, proteins are used as gelling agents, emulsifiers, foaming agents and thickeners. Many food proteins are enzymes which can enhance the rate of certain biochemical reactions. These reactions can have either a favorable or detrimental effect on the overall properties of foods. Food analysts are interested in knowing the total concentration, type, molecular structure and functional properties of the proteins in foods (Skjaerven et al., 2009).

MATERIALS AND METHODS

Experimental plants

Marjoram plants were obtained from Genetic Engineering and Biotechnology Research Institute (GEBRI), University of Sadat city, Sadat city, Egypt

Protein analysis

Discontinuous polyacrylamide gel electrophoresis (SDS-PAGE) was used for Total soluble protein analysis according to the method adopted by Stegman et al. (1987) and Hemeida (1994).

Sample preparation

Half gram from each sample of control and each treatment leaves was ground. The samples were extracted in 1ml of extraction buffer consisting of 0.15M Tris HCl, 2% Triton X100 and 1% ß-mercaptoethanol. The extracts were centrifuged at 15000 rpm for 30 minutes and the supernatant was stored at -20°C until used for electrophoresis.

For loading, 50μl of the extract was mixed with 25μl of SDS/mercaptoethanol (ME) solution (5% each) and incubated at 100°C for three min. in boiling water bath. Sucrose (10%) was added to the latter solution to increase the specific gravity of the supernatant. As a tracking dye, 0.01% Amido black (2μl of 0.5% Amido Black solution to 100μl of the supernatant) was added to the latter solution before use. Samples (30μl each) were then applied into the wells of the gel.

Stock solutions

Acrylamide monomer solution contained 30 gm Acryl amide, and 0.8 gm Bis- Acryl amide. Solution was brought up to 100 ml with water. Solution was filtered and stored at 4°C in dark (30 days max)
4.2. Separating gel buffer (1.5 M Tris-HCl pH 8.8) contained 18.168 gm Tris, and 20 ml 2N HCL. Solution was brought up to 100 ml with water. Solution was filtered and stored at 4°C in dark (30 days max)

4.3. Stacking gel buffer (1M Tris-HCl, pH 6.8) contained 12.11 gm Tris, and 17 ml 2N HCL. Solution was brought up to 100 ml with water. Solution was filtered and stored at 4°C in dark (30 days max)

4.4. SDS solution 10% (w/v) contained 10% SDS

4.5. Electrode buffer (0.05M Tris, 0.384 M glycine, 0.1% SDS pH 8.3) contained 28.8 gm Glycine, 8 gm Tris, and 1 gm SDS. Solution was brought up to 1 L with water.

4.6. Commassie blue contained 1 gm Commassie blue R-250, 0.25 gm Commassie blue G-250, 16 ml Methanol, 40 ml Acetic acid and 40 ml water.

4.7. Staining solution contained 60 gm TCA, 200 ml Methanol, 20 ml Acetic acid and 800 ml water.

4.8. Distaining solution contained 240 ml Methanol, 40 ml Acetic acid and 560 ml water.

Preparation of SDS polyacrylamide Gel

Discontinuous polyacrylamide gel consisted of a resolving or separating (lower) gel and stacking (upper) gel was used. The stacking gel acts to concentrate large sample volumes resulting in better band resolution. Gel was prepared according to Laemmli(1970)

5.1 Separating gel (10 %) contained 0.75 Monomer solution, 5 ml Separating buffer, 15 ml Water, 300 µl 10% SDS, 30µl TEMED and 75 µl Ammonium persulfate.

5.2. Preparation of stacking Gel (4%) contained 1.33 ml Monomer solution, 1.67 ml Stackingbuffer, 7 ml Water, 10.00 µl 10% SDS, 10.00 µl TEMED and 33.5 µl 10% Ammonium persulfate.

To prepare monomer solution of both separating and stacking gels, all reagents were combined, except TEMED and Ammonium persulfate which were added prior to casting the gels

Running conditions

Electrophoresis experiments (Shelton Scientific Mfg Gel electrophoresis apparatus) were conducted under cooling using a 150 volt electric current exerting its effect through the 6 hours running period.

Staining procedure

SDS polyacrylamide gels were kept overnight in the staining solution which was composed of 2.5 ml of 1% aqueous solution of Commassie Blue R-250 added to 100 ml TCA/Methanol/Acetic acid aqueous solution.

Distaining procedure

For distaining, the gels were immersed overnight in the distaining solution and then photographed and analyzed.
RESULTS

Protein analysis

This experiment was planned and achieved in order to determine any probable variation in the banding pattern of total soluble proteins due to the exposure of marjoram to pathogen, biocontrol agent, pathogen and single biocontrol agent or pathogen and combination treatment of biocontrol agents. Tissue extracts from leaves was assayed by electrophoretic analyses and then estimated by electropherograms scan unit (Pexil) and using Scannogram Phoretix 1 D software (Ibrahim et al., 2014)

Total soluble proteins patterns of leaves extracted from different treatments and control are shown in Figure 1. For control, healthy plants, plants and pathogen and plants and bioagent were used separately. Broadway Dual Pre-stained Protein Marker (in IRON Biotechnology, Germany) was used as molecular

Proteins bands recorded by the scannogram analysis are illustrated in Figure (2).

Figure 1. SDS electrophoresis of total soluble protein patterns of controls, different treatments and molecular marker. M: molecular marker, 1: Bacillus megaterium, 2: Fusarium aquaeductuum 3: Trichoderma harzinum + Fusarium aquaeductuum, 4: Control 5: Bacillus megaterium + Trichoderma harzinum + Trichoderma hamatum + Fusarium aquaeductuum.

Figure 2. Dendogram of total soluble protein patterns electrophoresis of controls, different treatments and molecular marker. M: molecular marker, 1: Bacillus megaterium, 2: Fusarium aquaeductuum, 3: Trichoderma harzinum, + Fusarium aquaeductuum, 4: Control, 5: Bacillus megaterium + Trichoderma harzinum + Trichoderma hamatum + Fusarium aquaeductuum

The Molecular weight of obtained bands and One/Zero data is presented in Table (1).
Table 1. Molecular weights of bands and One/Zero data for controls and different treatments.

<table>
<thead>
<tr>
<th>MW</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
<th>9</th>
</tr>
</thead>
<tbody>
<tr>
<td>40.361</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>35.88</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>32.369</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>28.296</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>24.221</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>19.963</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>16.839</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>12.547</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
</tbody>
</table>

MW: molecular weight, 1: *Bacillus megaterium*, 2: *Fusarium aquaeductuum* 4: *Trichoderma harzinum*, + *Fusarium aquaeductuum* 6 and 7: Control 8 and 9: *Bacillus megaterium* + *Trichoderma harzinum* + *Trichoderma hamatum* + *Fusarium aquaeductuum*.

DISCUSSION

Obtained results reveal that there is a total of 8 bands observed in the gel. The number of bands was the same (5 bands) for healthy plants and controls, healthy plants and biocontrol agent and diseased plants and combination treatment of biocontrol agents but with different molecular weights for some bands. With diseased plants and diseased plants and single biocontrol agent there were 7 and 6 bands, respectively.

Furthermore, the present data show that the specific protein molecular weights ranged from 40.361 to 12.547 KD (16). Bands number 2 (35.88 KD) and 3 (32.369 KD) were commonly observed in diseased plants, while band number 6 of molecular weight 19.963 KD was unique band detected in diseased plants and single biocontrol agent and in combination treatment of biocontrol agents. As observe in Figure (1) the density of band 6 in diseased plants and single biocontrol agent was more than that of diseased plants and combination treatment of biocontrol agents. There was a specific band (band number 8 with molecular weight of 12.547 KD) expressed in healthy plants and biocontrol agents, diseased plants and single biocontrol agent and diseased plants and combination treatment of biocontrol agents.
REFERENCES


